### A Mycotoxin Briefing

Written By: Steve Ensley, DVM, PhD, Kansas State Veterinary Diagnostic Laboratory Toxicologist Submitted By: Charlene Miller, Agriculture Agent KSRE/Butler County

Mycotoxins are secondary metabolites of fungi that are toxic to other life forms. More than 800 mycotoxins have been detected, but relatively few have been characterized and are considered to be important to animal health. There has been a tremendous increase in the ability to detect new and novel mycotoxins in the last few years.

Mycotoxin prevalence and concentration are sporadic and vary annually, even in the same location. Production is affected by local weather patterns, crop damage and productions practices. Mycotoxins may be produced preor post-harvest.

Mycotoxins known to occur in Kansa are aflatoxin, fumonisin, vomitoxin, T-2 and zearalenone. They are mostly found in grain, including corn, wheat, milo, rye, and oats. They may be found in silages or other feeds which contain grain.

#### Health effects:

Since there are many kinds of mycotoxins, which differ from each other, they produce many different kinds of diseases, called mycotoxicoses. They may cause diseases of the liver, gastrointestinal tract, respiratory system, immune system, reproductive system or kidneys. Mycotoxicoses are not contagious.

Effects of the mycotoxins known to occur in Kansas are described in the table below.

#### **Detection of mycotoxins:**

Detection of mycotoxins in feeds can aid in the prevention of mycotoxicoses.

Chemical analysis is the best way to find mycotoxins in feed. Screening methods such as ELISA kits are commonly used to rapidly detect mycotoxins. Positive results obtained by most screening methods do not prove the presence of mycotoxins, so positive results should be confirmed using chemical methods.

Screening for mycotoxins in feeds by black light (ultra-violet light) is not always reliable. Detection is based upon the fluorescence of chemicals indicative of the presence of aflatoxin, which occurs when the black light shines on them. Mycotoxins other than aflatoxin do not fluoresce. Chemicals that are not mycotoxins may also fluoresce, causing false positive results

### Sampling for analysis:

Mycotoxins are not evenly distributed in feeds. Some areas may contain very high concentrations; other areas may contain no detectable amounts. Detection in feeds depends upon the quality and quantity of sample provided for analysis. The sample should reflect all of the feed available at the time the problem occurred. The longer sample is collected after onset of animal health problems, the more likely the feed from which the sample is collected will not be characteristic of the feed being eaten at the time the problem began. And, the smaller the sample collected and provided for analysis, the more likely mycotoxin contamination will be missed.

Sampling based upon visible presence of molds does not always provide a sample that contains mycotoxins. The presence or absence of visible mold growth are not reliable indicators of the presence or absence of mycotoxins. Very moldy feed may not contain any detectable amounts of know mycotoxins, while good looking feed may contain very high concentrations.

It is best to collect a sample during movement of the feed, like when augering it from the storage bin into a grain truck. Collect small amounts over the entire time the feed is being moved, so that at least 5 pounds, and as much as 10 pounds, have been collected, and mix the collected sample thoroughly. From this large sample randomly select 1 pound in a zip lock bag to submit to the laboratory. Probe sampling may be used. Collect probe samples from as many areas of the feed as practical into a composite sample to select the 1 pound sample from.

Dry samples are preferred for transport to the laboratory. Mold may grow on wet sample, especially if the sample is placed in a plastic bag. Oven-dry specimens to less than 13% moisture for best preservation. Ship sample in a Styrofoam container on ice.

If you have questions about Mycotoxins or other potential poisons, please contact Kansas State Veterinary Diagnostic Laboratory Outreach at 866-512-5650 or clientcare@vet.k-state.edu

# Table. Use of mycotoxin-contaminated feeds:

There are legal requirements in regard to mixing or blending mycotoxin contaminated feeds.

## MYCOTOXINS COMMONLY FOUND IN KANSAS GRAINS AND THEIR HEALTH EFFECTS

Mycotoxin <b>Source</b>		Substances		Conditions favoring production	Effects		Remarks	
<b>Aflatoxins</b> (B <sub>1</sub> , B <sub>2</sub> , G <sub>1</sub> , G <sub>2</sub> )	Aspergillu flavus, Aspergillu parasiticu	us corn, milo, us us seed, peanuts		D,	78°F - 90°F (ideal) down to 55°F Kernel damage High humidity or grain moisture	Acute: liver toxin, evidence of liver pathology including depression and anorexia Chronic: depends upon species, but includes anorexia, poor growth, anemia, ascites, steatorrhea		The first mycotoxin characterized. More of a problem in states SE of Iowa, especially in the SE part of the US. Chronic exposure can produce liver cancer
Fusarium moniliforme, Fusarium proliferatum		White & yellow corn		Not well defined: Drought during growing season, followed by cool, wet conditions during pollination and development			Horses: equine leukoencephalomalacia (ELE) Swine: respiratory syndrome, icterus, weight loss, reduced feed intake Ruminants: anorexia and mild weight loss on diets with up to 200 ppm fumonisins, no deaths, no other significant or persistent signs Poultry: when consuming feed containing 200 - 400 ppm may develop inappetence and skeletal	

				abnorm than ma	alities. More resistant ammals
<b>Vomitoxin</b> (deoxynivalenol, DON)	Fusarium roseum	Corn, milo, wheat, rye, barley, other cereal crops	Alternating cool and warm temperatures Wet period during flowering	Swine: associated with feed refusal, resulting in decreased weight gain Cattle: no apparent adverse health effects	Often occurs with zearalenone
Zearalenone	Fusarium roseum (Fusarium graminearum), Fusarium moniliforme	Corn, wheat, barley, milo, oats	High moisture content (> 22%) Alternating high and low temperatures during the maturing and harvesting stage (45°F - 70°F)	Functions as a weak estrogen. It does not cause abortions. Swine: females, hyperestrogenism, nymphomania; anestrus, pseudopregnancy; immature males, feminization, reduced libido, retarded testicular development; mature males, no effects < 200 ppm Cattle: little effects; vaginal secretions, vaginitis, mammary enlargement	Mold infection commonly called "pink ear rot" or "scab" Often occurs with DON (deoxynivalenol, vomitoxin)

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